

Mini Review

Bioactivity studies and chemical constituents of *Murraya paniculata* (Linn) Jack

¹ Ng, M. K., ¹Abdulahadi-Noaman, Y., ²Cheah, Y.K., ¹Yeap, S. K. and
^{1*}Alitheen, N.B.

¹Department of Cell and Molecular Biology
Faculty of Biotechnology and Biomolecular Sciences
Universiti Putra Malaysia, 43400 Serdang, Selangor, Malaysia

²Unit of Molecular Biology and Bioinformatics
Department of Biomedical Science
Faculty of Medicine and Health Sciences
Universiti Putra Malaysia, 43400 Serdang, Selangor, Malaysia

Article history

Received: 4 January 2011
Received in revised form:
6 April 2012
Accepted: 6 April 2012

Keywords

Murraya paniculata,
“kemuning”,
folk medicines,
bioactivities,
chemical constituents

Abstract

Murraya paniculata (Linn) Jack (Orange Jasmine), known as “Kemuning Putih” in Malaysia, has been widely used as food flavor additive in cuisine by local residences. This is due to the strong fragrances of the leaves which make it suitable to be used in Indian and Malay dishes. Besides as a flavoring, leaves, branches, stem barks and roots of the plant are used in folk medicine to treat dysentery and morning sickness. Flowers of the plants are used in cosmetics. Since 1970’s, flavonoids and coumarins were isolated from *Murraya paniculata*, but no further bioactivity has been tested from the isolated compounds. The aim of this paper is to review and update the research related to chemical constituents and bioactivities of *Murraya paniculata* (L) Jack.

© All Rights Reserved

Introduction

Asia has been well known as “Land of spices” since ancient where places like Maluku Island, Indonesia (Spices Island), Sumatera (Spice Isle) and Melaka (Spice City) were popular as the spice markets. In ancient, people not merely use spices to add flavour to foods and beverages, but also as medicines, disinfectants, incenses, stimulants and aphrodisiac agents. Spices were used to cure, relax and excite human being and greatly influence their daily life (Chomchalow, 1996).

Spices are known as non-leafy parts such as bud, fruit, seed, bark, rhizome and bulb of plants that as flavoring or seasoning where some of them may also used as herb medicinal. While the similar phrase “herb” used to distinguish the same plant parts which derived from leafy or soft flowering (Chomchalow, 1996). In Malaysia, one of the most commonly used herbs in Malaysia cuisines is *Murraya paniculata* which is commonly known as “kemuning putih”. Malaysians commonly use *M. paniculata* leaves in preparing soup, fish and meat. Recently, it has also been used to prepare spicy chicken dishes in one of the most popular fast food restaurant in Malaysia.

Among the 14 species under the genus of *Murraya*, *Murraya paniculata* (Linn) Jack and *Murraya koenigii* (L) Spreng are the most popular natural flavor with

the potential bioactivities. Since both are closely related species, they were named as “curry leaves” in Malaysia. However, *M. paniculata* is often found in Peninsular Malaysia (Kedah, Pulau Pinang, Kelantan, Perak, Melaka, Johor) and Singapore (Table 1), whilst *M. koenigii* was grown natively in Thailand (Table 1). *M. paniculata* is an 8 to 12 feet high medium-sized shrub with white, fragrance flowers blooming all the year round and the leaves are green obovated in shape, with less than 2 inches of blade length, alternately arranged on the branch (Gilman, 1999). Another widespread species, *Murraya exotica* which was previously thought as synonym with *M. paniculata* has smaller, obovated leaflets compare to

Table 1. Differences between *M. paniculata*, *M. koenigii* and *M. exotica* (Gilman, 1999)

	<i>Murraya paniculata</i> (L) Jack	<i>Murraya koenigii</i> (L) Spreng	<i>Murraya exotica</i> (L)
Leaves	Pinnate, with 3-7 leaflets	Pinnate, with 12-23 leaflets	Foliate, with 3-7 short leaflets
Flowers	Whitefish	Small, white, fragrance	White oblong, fragrance
Fruits	Small berry dotted with oil cells, bright red in colour	small black berries	Not known
Habitat	Peninsular Malaysia and Singapore	Grown native in Thailand	Widespread in tropical and subtropical areas
Local name	“Kemuning putih”	“Kari”	Previously known as Kemuning putih

*Corresponding author.

Email: noorjahan@biotech.upm.edu.my

M. paniculata. Taxonomic divergent has then resolved by using cumulative data for three Single Primer Amplification Reaction methods (ITS, RAPD and AMD), to compute pair-wise distances, where wide range in distances reported between *M. paniculata* and *M. exotica*. This also has been congruent through comparing the morphological findings suggesting both plants came from different taxa (Verma *et al.*, 2009).

Due to the rapid development of Western therapeutic, interest in applying medicinal plants were faded. However, the awareness of potential source of new drug discoveries from plant world rose past few years. Researches included crude extract isolation, bioactivities studies and also structure elucidation of isolated compounds. Herbal medicines are being emphasised with their ready availability and minimal side effects compared to synthetic drugs (Ho *et al.*, 2009; Yeap *et al.*, 2010). In many cases, crude drugs are found more potent than synthetic drugs on the biological activity with lower toxic effect because of the synergistic effect of other presented compounds. Since *M. paniculata* are not well studied as *M. koenigii*, this review was aimed to give an overview on the current studies on the bioactivities of *M. paniculata* and thus gives some insights into future research and commercial value of this plant.

Activities of *Murraya paniculata* (Linn) Jack

“Kemuning” is a plant under the family of Rutaceae where an anti-implantation agent named yuehchukene was isolated (Kong *et al.*, 1986). With a single dosage or single day of 3 mg/ kg on 2nd day pregnancy rats after successful mating, Yuehchukene extracted from roots of *M. paniculata* was reported 100% active in anti-fertility ability (Kong *et al.*, 1985). In one chemotaxonomic research, this yuehchukene is potentially found in *Murraya* species members especially *M. exotica* (2.61 mg/100 g), *M. alata* (2.06 mg/100 g) and *M. paniculata* (1.39 mg/100 g) (Kong *et al.*, 1986). Besides, *M. paniculata* was also reported to have antinociceptive effect and toxicity effect on the ethanolic leaf extract. Writhing inhibition 26.67% and 66.67% were reported at 250 mg/ kg and 500 mg/ kg of body weight mice while toxicity towards brine shrimp was determined at LD₅₀ 32 µg/mL in the same study (Sharker *et al.*, 2009).

Antioxidant activity

Currently, *M. paniculata* with several extraction methods possess antioxidant activity. According to Rohman and Riyanto (2005), ethanolic extract of “kemuning” leaves using linoleic-thiocyanate method showed antioxidant strength in the following

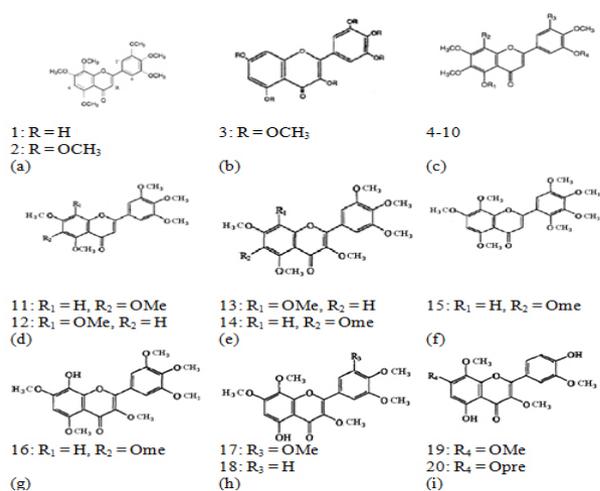


Figure 1. Chemical structures of flavonoids isolated from different parts of *M. paniculata*. (Source: Figure 1(a) Silva *et al.*, 1980 and Yang and Du, 1984; Figure 1(b) Wu *et al.*, 1993 ; Figure 1(c) Kinoshita and Firman,1995; Figure 1(d)-(i) Ferracin *et al.*, 1998)

sequence of 10% “kemuning” > 1% vitamin E > 5% “kemuning” > 1% of “kemuning” extract. Using 2,2-diphenyl-1-picryl hydrazyl (DPPH) method, the IC₅₀ of “kemuning” extract was 126.17 µg/ mL which is 15 times lower than the vitamin E (positive control) 8.27 µg/ mL. In addition, the acetone extraction of *M. paniculata* showed inhibitory effect toward xanthine oxidase (XO), tyrosinase and lipoxygenase (LOX) where 100 µg/ mL of the acetone extract was able to inhibit 10% of XO activity, 62% of LOX activity and at 500 µg/mL, the acetone extract inhibited 72% of tyrosinase activity (Chen *et al.*, 2009).

Antimicrobial activity

According to Aziz *et al.*, (2010), auraptene, trans-gleinadiene, 5,7-dimethoxy-8-(3-methyl-2-oxo-butyl) coumarin and toddalenone were isolated from the chloroform, petroleum ether and methanol extract from leaves of *M. paniculata*. Only chloroform extract showed a weak activity against *Bacillus cereus* and *Saccharomyces cerevisiae* with inhibition zone 9 mm and 8 mm respectively. Among the isolated compound, only trans-gleinadiene exhibited a weak antimicrobial activity against *Bacillus cereus* with 8 mm inhibition zone which conclude that trans-gleinadiene, auraptene and 5,7-dimethoxy-8-(3-methyl-2-oxo-butyl) coumarin give synergistic effect towards chloroform extract.

Constituents in *Murraya paniculata* (Linn) Jack Flavonoids

Methanolic extract of *M. paniculata* (Jack) leaves was found to contain 3',4',5,5',7,8-hexamethoxyflavone (Figure 1a) and 3,3',4',5,5',7,8-heptamethoxyflavone (Figure 1a) (Silva *et al.*, 1980; Yang and Du, 1984). From the fresh flower

Table 3. Three indole alkaloids isolated from leaves, root bark and fresh flowers of *M. paniculata* (Kong et al., 1986; Wu et al., 1989; Wu et al., 1993)

No.	Indole alkaloids	Mol. Formula	Source	Substituents
				R
1	Yuehchukene (Figure 2a)	C ₂₆ H ₂₆ N ₂	Leaves	-
2	Murrayacarine (Figure 2b)	C ₁₄ H ₁₃ NO ₃	Root bark	-
3	Murrayaculatin (Figure 2c)	C ₁₀ H ₉ NO ₃	Fresh flower	-

(Figure 2). While methyl 2,5-dihydroxycinnamate and murrayatin (Rahman et al., 1997) were reported in the methanolic extract of leaves. Cytotoxic effect of water extracts from leaves and branches of *Philadelphus coronaries* L. (Hydrangeaceae) was investigated on A431 (human skin carcinoma cell line) and MCF-7 (human breast adenocarcinoma cell line). Both leaves and branches extracts gave ED₅₀ = 2.19 µg/mL on MCF-7 and ED₅₀ = 27.95 µg/mL on A431 at 24 hours incubation period (Valko et al., 2006). This cytotoxic effect on A431 and MCF-7 may due to the presences of coumarins (umbelliferone and scopolin) which also found in *M. paniculata* extract.

Compounds in *Murraya paniculata*

Composition of *Murraya paniculata* essential oil:

Besides alkaloids, flavonoids and coumarins, leaves of *M. paniculata* also contained 60 compounds (Table 5) being identified from volatile and essential oil extracted from the leaves. The major components were γ-elemene (31.7%), perolidol (10%), t-caryophyllene (11.6%), caryophyllene oxide (16.6%), β-caryophyllene (11.8%), spathulenol (10.2%), β-elemene (8.9%), germacrene D (6.9%) and cyclooctene, 4-methylene-6-(1-propenylidene) (6.4%) (Chowdhury et al. 2008; Li et al., 1988; Rout et al., 2007). According to Chowdhury et al. (2008), 58 compounds were found from the oil of *M. paniculata*. While the major compounds found are (E)-caryophyllene (Table 5) was found to possess cytotoxic against MDA-MB-231 (IC₅₀ = 31.6 µg/mL) and Hs 578T (IC₅₀ = 78.3 g/mL) human tumor cells (Palazzo et al., 2009). Besides, oils extracted from leaves and berries of *Juniperus phoenicea* show similar cytotoxic activity on U251 (0.6 µg/ml), HeLa (5.0 µg/ml) but slightly higher of berry oil compared to leaves against H460 (0.6 and 0.7 µ/ml, respectively), HepG2 (0.7 and 0.9 µg/ml, respectively), MCF-7 (0.8 and 1. µg/ml, respectively) cell lines. These berries and leaf oil from *Juniperus phoenicea* are rich with Monoterpene hydrocarbons (El-Sawi et al., 2007) which are also found in *M. paniculata*.

Polysaccharide and others

A water soluble gum polysaccharide was isolated from the fruit of *M. paniculata* (Mondal et al.,

Table 4. Coumarins (Wu et al., 1989) isolated from the root bark, fresh flowers of *M. paniculata* (Lin and Wu, 1994; Kinoshita et al., 1996; Rahman et al., 1997)

No.	Coumarin/Compounds	Mol. Formula	Source	Substituents R
1	3-formyl indole	C ₉ H ₇ O	Root bark	-
2	omphalo-car-Pn	C ₁₇ H ₂₂ O ₆		-
3	5,7-dimethoxy-8-(3'-methoxy-2'-oxobutyl)-coumarin	C ₁₆ H ₁₈ O ₅		-
4	coumurrayin	C ₁₆ H ₁₈ O ₄		-
5	murrayatin	C ₁₆ H ₂₀ O ₆		-
6	omphalin	C ₁₂ H ₁₆ O ₁₆		-
7	murrayol	C ₁₅ H ₁₆ O ₄		-
8	(-)-murrayarin	C ₁₆ H ₁₈ O ₅		-
9	(k)-murrayarin	C ₁₆ H ₁₈ O ₅		-
10	murrayidin	C ₁₅ H ₁₆ O ₅		-
11	mexotian	C ₁₆ H ₂₀ O ₆		-
12	murrayatin	C ₁₅ H ₁₆ O ₅		-
13	ferulyl esters	C ₁₅ H ₁₆ O ₅		-
14	yuehchin A	C ₁₆ H ₁₈ O ₅	Fresh flowers	CH ₂ CH(OH)C(CH ₃)OOC(CH ₃)OH
15	yuehchin B	C ₁₆ H ₂₀ O ₅		CH ₂ CH(OH)C(CH ₃)OCH ₃
16	yuehchin C	C ₁₇ H ₂₀ O ₅		CH ₂ CH(OH)C(CH ₃)OCH ₂ CH ₃
17	murrayarin	C ₁₆ H ₁₈ O ₅		C(OCH ₃)H-C(OH)H(C(CH ₃)=CH ₂)
18	murrayidin	C ₁₅ H ₁₆ O ₅		C(OCH ₃)H-C(OH)HCC(CH ₃)=CH ₂
19	isomeranzin	C ₁₅ H ₁₆ O ₅		CH ₂ COCH(CH ₃) ₂
20	murrayolin	C ₁₅ H ₁₆ O ₅		C(COH)=C(CH ₃) ₂
21	7-methoxy-8-(4'-ethoxy-2'-methoxybutyl)-coumarin	Not known		CH(OAc)COCH(CH ₃) ₂
22	murrayolin	Not known		CH(OH)COCH(CH ₃) ₂
23	Scopolin	C ₁₀ H ₁₄ O ₄		H
24	7-methoxy-8-(4'-ethoxy-3'-methoxybutyl)-coumarin	Not known		CH(OEt)CH(OH)C(CH ₃)=CH ₂
25	Umbelliferone	Not known		-
26	Paniculatin	C ₂₇ H ₃₆ O ₁₅		CH(O)COCH ₂ CH(CH ₃) ₂ COCH(CH ₃) ₂
27	Braylin	C ₁₅ H ₁₈ O ₄		-
28	Auraptenol	Not known		CH ₂ C(OH)C(CH ₃)=CH ₂
29	Meranzin hydrate	C ₁₅ H ₁₈ O ₅		CH ₂ C(OH)C(CH ₃) ₂ OH
30	Murrayidin	C ₁₅ H ₁₆ O ₅		C(OH)H-C(OH)H-C(CH ₃)=CH ₂
31	Scopolin	C ₁₀ H ₁₄ O ₄	Glucose	
32	Califen	C ₈ H ₁₀ N ₂ O ₂	-	
33	3,3',4',5',6',7'-hexamethoxy-flavone	Not known	-	
34	4-hydroxybenzylide	C ₇ H ₈ O ₂	-	
35	p-hydroxybenzoic acid	C ₇ H ₆ O ₃	-	
36	Cis-fenolic acid and transferulic acid	Not known	-	
37	Cis-methyl ferulate and transferulic acid	C ₁₁ H ₁₂ O ₄	-	
38	Trans-ethyl ferulate	Not known	-	
39	Murrayolin	C ₁₅ H ₁₆ O ₅	Leaves	-
40	Isomurrayolin and isovalerate	Not known		-
41	Murrayatin	C ₁₅ H ₁₆ O ₅		-
42	Minu microin	C ₁₄ H ₁₆ O ₄		-
43	Toddalenone	C ₁₅ H ₁₈ O ₅		-
44	Omphalourayin	Not known		-
45	Coumurrayin	C ₁₅ H ₁₆ O ₄		-
46	Auraptenone	C ₁₀ H ₁₄ O ₄		-
47	Toddalin	Not known		-
48	methyl 2-methoxy-5-hydroxycinnamate	C ₁₁ H ₁₂ O ₄		CH ₃
49	methyl 2,5-dihydroxycinnamate	C ₁₀ H ₁₀ O ₄		H
50	8-ethoxy-3-methylbutoxy-7-methoxy coumarin	C ₁₅ H ₁₈ O ₅		-
51	murrayatin	C ₂₀ H ₂₀ O ₆	-	

Table 5. Composition of volatile oils (Li et al., 1988) and essential oil (Rout et al., 2007; Chowdhury et al., 2008) isolated from leaves of *M. paniculata*

No.	Component	Mol. Formula	No.	Component	Mol. Formula
1	δ-Elemene	C ₁₅ H ₂₄	64	Ethyl palmitate	C ₁₈ H ₃₆ O ₂
2	β-Elemene	C ₁₅ H ₂₄	65	Palmitic acid	C ₁₈ H ₃₆ O ₂
3	1-Caryophyllene	C ₁₅ H ₂₄	66	Mono oil	C ₁₈ H ₃₆ O
4	1-β-Farnesene	C ₁₅ H ₂₄	67	Methyl indole	C ₁₀ H ₈ O
5	Eranopholene	C ₁₅ H ₂₄	68	Methyl indole	C ₁₀ H ₈ O
6	Humulene	C ₁₅ H ₂₄	69	Methyl stearate	C ₁₈ H ₃₆ O ₂
7	alloaromadendene	C ₁₅ H ₂₄	70	9,12-Octadecanoid	C ₁₈ H ₃₆ O
8	β-Cubebene	C ₁₅ H ₂₄	71	Dogonane	C ₂₂ H ₃₆
9	α-Bergamotene	C ₁₅ H ₂₄	72	Sabinene	C ₁₀ H ₁₆
10	γ-Elemene	C ₁₅ H ₂₄	73	3-Hexen-1-d, formate	C ₈ H ₁₆ O ₂
11	δ-Cadinene	C ₁₅ H ₂₄	74	Limonene	C ₁₀ H ₁₆
12	Pero linalol	Not known	75	Linalol	C ₁₀ H ₁₆ O
13	Nonotaton	C ₁₅ H ₂₄ O	76	Cyclohexenyl-4-ethyl-3-methyl	C ₁₀ H ₁₆
14	Toreol	C ₁₅ H ₂₄ O	77	Cyclohexenyl-6-ethyl-3-methyl	C ₁₀ H ₁₆
15	Bulnesol	C ₁₅ H ₂₄ O	78	Azulen	C ₁₀ H ₁₆
16	Benzaldehyde	C ₇ H ₆ O	79	Ocimen	C ₁₀ H ₁₆
17	Myrcene	C ₁₀ H ₁₆	80	cis-3-Hexenylvalerate	C ₁₀ H ₁₈ O ₂
18	Limonene	C ₁₀ H ₁₆	81	2-Geranyl-1-ene-2-methyl-5-(1-methyl-ethyl)	C ₁₈ H ₃₀
19	(Z)-β-Ocimene	C ₁₀ H ₁₆	82	1H-Imidazole-4-methyl-5-methyl	Not known
20	(E)-β-Ocimene	C ₁₀ H ₁₆	83	α-Cubebene	C ₁₅ H ₂₄
21	α-Terpinene	C ₁₀ H ₁₆	84	3,9-Dodecadiene	C ₁₂ H ₂₂
22	Methyl butyrate	C ₈ H ₁₆ O ₂	85	β-Caryophyllene	C ₁₅ H ₂₄
23	Linalol	C ₁₀ H ₁₆ O	86	Caryophyllene oxide	C ₁₅ H ₂₄ O
24	Methyl phenylacetate	C ₁₀ H ₁₀ O ₂ C ₆ H ₅ CH ₂ COO CH ₃	87	Cyclohexenyl-3-methyl-2-(1-propenyl)	C ₁₅ H ₂₄
25	Methyl salicylate	C ₉ H ₁₀ O ₂	88	Retinal	C ₂₀ H ₃₀ O
26	Nerol	C ₁₀ H ₁₆ O	89	α-Caryophyllene	C ₁₅ H ₂₄
27	2-Phenylethylacetate	C ₁₂ H ₁₄ O ₂	90	β-Humulene	C ₁₅ H ₂₄
28	Indole	C ₈ H ₇ N	91	Copaene	C ₁₅ H ₂₄
29	α-Cubebene	C ₁₅ H ₂₄	92	Cubanol	C ₁₅ H ₂₄ O
30	Cyclosativene	C ₁₅ H ₂₄	93	α-Bulnesene	C ₁₅ H ₂₄
31	α-Copaene	C ₁₅ H ₂₄	94	Calamene	C ₁₅ H ₂₄
32	Phenethylisobutyrate	C ₁₂ H ₁₆ O ₂	95	3-Tetradecenoic acid	C ₁₄ H ₂₆ O ₂
33	(Z)-Jasmone	C ₁₁ H ₁₆ O	96	Linalol, cis	C ₁₀ H ₁₆ O
34	(E)-Caryophyllene	C ₁₅ H ₂₄	97	β-Vetivene	Not known
35	Clovene	C ₁₅ H ₂₄	98	Nerolidyl acetate	C ₁₇ H ₂₈ O ₂
36	(Z)-β-Farnesene	C ₁₅ H ₂₄	99	Alloaromadendene oxide	C ₁₅ H ₂₄ O
37	α-Humulene	C ₁₅ H ₂₄	100	Spathulol	C ₁₅ H ₂₄ O
38	(E)-β-Farnesene	C ₁₅ H ₂₄	101	D-Verbenone	C ₁₀ H ₁₆ O
38	Murola-4(14)5-diene	C ₁₅ H ₂₄	102	Pyrimidinol(4H)thione, 3,4-dihydro-6-methyl,4-phenyl	C ₈ H ₈ N ₂ S
40	Germacren D	C ₁₅ H ₂₄	103	3-Carene	C ₁₀ H ₁₆
41	ar-Curumen	C ₁₅ H ₂₄	104	12-Oxobicyclo(9,1,0)decane-3,7-dienel 5,5,8-tetramethyl	C ₁₈ H ₃₀ O
42	Bicydogermacrene	C ₁₅ H ₂₄	105	Globulol	C ₁₅ H ₂₄ O
43	α-Zingiberene	C ₁₅ H ₂₄	106	Eranopholene	C ₁₅ H ₂₄
44	(E,E)-α-Farnesene	C ₁₅ H ₂₄	107	2(4H)-Npithalene, 4,4,6,6,7,8,8-hexahydro,4,8-dimethyl	C ₁₀ H ₁₆ O
45	β-Curumen	C ₁₅ H ₂₄	108	Tai-Muobol	C ₁₅ H ₂₄ O
46	δ-Cadinene	C ₁₅ H ₂₄	109	Leolol	C ₁₅ H ₂₄ O
47	Cadinal-4-diene	C ₁₅ H ₂₄	110	Aromadendene oxide	C ₁₅ H ₂₄ O
48	Germacrene B	C ₁₅ H ₂₄	111	α-Calamene	Not known
49	(E)-Nerolid	C ₁₅ H ₂₄ O	112	Longifidenealdehyde	C ₁₅ H ₂₄ O
50	Spathulol	C ₁₅ H ₂₄ O	113	11-Hexadecylol	C ₁₆ H ₃₄ O
51	Globulol	C ₁₅ H ₂₄ O	114	Cyclohexenyl-3-methyl-2-hydroxy-8-	C ₁₅ H ₂₄ O
52	Phenylethyl acetate	C ₁₂ H ₁₄ O ₂	115	Longipinocaradiol, tens	C ₁₅ H ₂₄ O
53	1-epi-Cubanol	C ₁₅ H ₂₄ O	116	Carvool	C ₁₀ H ₁₆ O
54	epi-α-Murrid	C ₁₅ H ₂₄ O	117	1-(3-cyclohexenyl-2-methyl)-2,6,6-trimethyl	Not known
55	α-Cadinol	C ₁₅ H ₂₄ O	118	1-Methylverbanol	C ₁₁ H ₁₆ O
56	Patchouliolalcohol	C ₁₅ H ₂₄ O	119	Cyclopropane-1,6-dimethyl-2,2,3,3-tetramethyl-propyl-1-ynyl	C ₁₀ H ₁₆ Br
57	β-Bisabolol	C ₁₅ H ₂₄ O	120	Corymbolone	C ₁₅ H ₂₄ O
58	Benzyl butyrate	C ₁₂ H ₁₆ O ₂	121	2(4a,8-Dimethyl, 3,3,4,4,6,6-tetraoxabicyclo[2.2.1]hept-2-en-1-yl)-prop-2-en-1-ol	Not known
59	Phenethylbutyrate	C ₁₂ H ₁₆ O ₂	122	1,2-dimethyl-3-hydroxy-4-oxo-2-pentene	C ₁₅ H ₂₄ O
60	Benzyl salicylate	C ₁₂ H ₁₄ O ₂	123	Anistolene oxide	C ₁₅ H ₂₄ O
61	Methyl palmitate	C ₁₇ H ₃₄ O ₂	124	6-(propyl-1,8-dimethyl-1,2,3,5,6,7,8,8a-octahydro-naphthalen-2-yl)	Not known
62	Phytol	C ₂₀ H ₄₀ O	125	Longifidene[2]-epoxide	C ₁₅ H ₂₄ O
63	Phenylethyl salicylate	C ₁₄ H ₁₆ O ₂			

2001). The polysaccharide have 1,3-linked β-D-galactopyranosyl, 1,3,6-linked β-D-galactopyranosyl, terminal β-D-galactopyranosyl and terminal α-D-galactopyranosyl, 1,4-β-D-galactopyranosyl. Although plenty of compounds were isolated from this plant, but further bioactivities has not been widely investigated yet.

Conclusion

In short, *Murraya paniculata* is rich of various types of active components. However, the bioactivities studies such as antioxidant, antimicrobial, anticancer, anti-diabetic and others have yet to be discovered. This plant should be given more attention since it could be easily obtained in Peninsular Malaysia.

References

Azizi, S.S.S.A., Sukari, M.A., Rahmani, M., Kitajima, M. and Ahpandi, N.J. 2010. Coumarins from *Murraya paniculata* (Rutaceae). The Malaysian Journal of Analytical Sciences 14:1-5.

Chen, C.H., Chan, H.C., Chu, Y.T., Ho, H.Y., Chen, P.Y., Lee, T.H. and Lee, C.K. 2009. Antioxidant activity of some plant extracts towards xanthine oxidase, lipoxygenase and tyrosinase. Molecules 14: 2947-2958.

Chomchalow, N. 1996. Spice Production in Asia – An Overview.

Chowdhury, J.U., Bhuiyan, M.N.I. and Yusuf, M. 2008. Chemical composition of the leaf essential oils of *Murraya koenigii* (L.) Spreng and *Murraya paniculata* (L.) Jack. Bangladesh Journal Pharmacological Society 3: 59-63.

El-Sawi, S.A., Motawae, H.M. and Ali, A.M. 2007. Chemical Composition, Cytotoxic Activity and Antimicrobial Activity of Essential Oils Of Leaves and Berries of *Juniperus phoeniceal* Grown In Egypt. African Journal of Traditional, Complementary and Alternative Medicines 4: 417-426.

Ferracin, R.J., Silva, M.F., Fernandes, J.B. and Vieira, P.C. 1998. Flavonoids from the fruits of *Murraya paniculata*. Phytochemistry 47: 393-396.

Gilman E.F. 1999. *Murraya paniculata*. Fact Sheet FPS 416 University of Florida.

Ho, W.Y., Ky, H., Yeap, S.K., Raha, A.R., Omar, A.R., Ho, C.L. and Alitheen, N.B. 2009. Traditional practice, bioactivities and commercialization potential of *Elephantopus scaber* Linn. Journal of Medicinal Plants Research 3: 1212-1221.

Ito, C., Itoigawa, M., Nakao, K., Murata, T., Tsuboi, M., Kaneda, N. and Furukawa, H. 2006. Induction of apoptosis by carbazole alkaloids isolated from *Murraya koenigii*. Phytomedicine 13: 359-365.

Kinoshita, T. and Firman, K. 1996. Highly oxygenated flavonoids from *Murraya paniculata*. Phytochemistry 42: 1207-1210.

- Kinoshita, T., Wu, J.B. and Ho, F.C. 1996. The isolation of a prenylcoumarin of chemotaxonomic significance from *Murraya paniculata* var. *omphalocarpa*. *Phytochemistry* 43: 125-128.
- Kong, Y.C., Ng, K.H., Wat, C.K.H., Wong, A., Saxena, L.F., Cheng, K.F., But, P.P.H. and Chang, H.T. 1985. Yuehchukene - a novel anti-implantation indole alkaloid from *Murraya paniculata*. *Planta Medica* 49: 304-307.
- Kong, Y.C., Ng, K.H., But, P.P.H., Li, Q., Yu, S.X., Zhang, H.T., Cheng, K.F., Soejarto, D.D., Kan, W.S. and Waterman P.G. 1986. Sources of the anti-implantation alkaloid yuehchukene in the genus *Murraya*. *Journal of Ethnopharmacology* 15: 195-200.
- Li, Q., Zhu, L.F., But, P.P.H., Kong, Y.C., Chang, H.T. and Waterman, P.G., 1988. Monoterpene and Sesquiterpene rich oils from the leaves of *Murraya* Species: Chemotaxonomic Significance. *Biochemical Systematics and Ecology* 16: 491-494.
- Lin, J.K. and Wu, T.S. 1994. Constituents of flowers of *Murraya paniculata*. *Journal of the Chinese Chemical Society* 41: 213-216.
- Mondal, S.K., Ray, B., Ghosal, P.K., Telean, A. and Vuorinen, T. 2001. Structural features of a water soluble gum polysaccharide from *Murraya paniculata* fruits. *International Journal of Biological Macromolecules* 29: 169-174.
- Palazzo, M.C., Agius, B.R., Wright, B.S., Haber, W.A., Moriarity, D.M. and Setzer, W.N., 2009. Chemical Compositions and Cytotoxic Activities of Leaf Essential Oils of Four Lauraceae Tree Species from Monteverde, Costa Rica. *Record of Nature Products* 3: 32-37.
- Parmar, C. and Kaushal, M. K. 1982. *Murraya koenigii*. In Parmar, C. and Kaushal, M.K (eds). *Wild Fruits*. p.45 – 48. India: Kalyani .
- Rahman, A. U., Sharbbir, M., Sultan, S. S. Z, Jabbar, A. and Choudhary, M.I. 1997. Cinnamates and coumarins from the leaves of *Murraya paniculata*. *Phytochemistry* 44: 683-685.
- Rohman, A. and Sugeng, R. 2005. Antioxidant potency of ethanolic extract of Kemuning leaves (*Murraya paniculata* (L) Jack) *in vitro*. *Majalah Farmasi Indonesia*, 16: 136-140.
- Rout, P.K., Rao, Y.R., Sree, A. and Naik, S.N. 2007. Composition of essential oil, concrete, absolute, wax and headspace volatiles of *Murraya paniculata* (Linn.) Jack flowers. *Flavour And Fragrance Journal* 22: 352-357.
- Sharker, S.Md., Shahid, I.J. and Hasanuzzaman, Md., 2009. Antinociceptive and bioactivity of leaves of *Murraya paniculata* (L.) Jack, *Rutaceae*. *Brazilian Journal of Pharmacognosy* 19: 746-748.
- Silva, L.B., Silva, U. L. L., Mahendran, M. and Jennings, R.C., 1980. Flavonoids of *Murraya paniculata* (Linn.) Jack. *Journal of the National Science Council of Sri Lanka* 8: 123-125.
- Valko, V., Fickova, M., Pravdova, E., Nagy, M., Grancai, D. and Czigle, S. 2006. Cytotoxicity of Water Extracts from Leaves and Branches of *Philadelphus coronarius* L. *Biomedical papers of Medical Faculty of the University Palacky Olomouc Czech Republic* 150: 71-73.
- Verma, S., Rana, T. S. and Ranade, S. A. 2009. Genetic variation and clustering in *Murraya paniculata* complex as revealed by single primer amplification reaction methods. *Current Science* 96: 1210-1216.
- Wu, T.S., Chan, Y.Y., Leu, Y.L. and Huang, S.C. 1994. A flavonoid and indole alkaloid from flowers of *Murraya paniculata*. *Phytochemistry* 37: 287 -288.
- Wu, T.S., Lin, C.N., Yang, L.K. and Lin, S.T. 1974. Studies of the constituents of *Murraya paniculata* Jack (L). *EJ52-1975-163*.
- Wu, T.S., Liou, M.J., Jong, T.T., Chen, Y.J. and Lai, J.S. 1989. Indole alkaloids and coumarins from the root bark of *Murraya paniculata* var. *omphalocarpa*. *Phytochemistry* 28: 2873-2874.
- Yang, J.S. and Du, M.H. 1984. Studies on the constituents of *Murraya paniculata* (L.) Jack grown in Yunnan. *Acta Botanica Sinica* 26: 184-188.
- Yeap, S.K., Ho, W.Y., Beh, B.K., Liang, W.S., Ky, H., Yousr, A.H.N. and Alitheen, N.B. 2010. *Vernonia amygdalina*, an ethnoveterinary and ethnomedical used green vegetable with multiple bio-activities. *Journal of Medicinal Plants Research* 4: 2787-2812.